Modified Atmosphere Packaging (MAP): Microbial Control and Quality

Introduction

Manipulation of the food product microenvironment using combinations of gaseous atmospheres has been in practice since the early 1930s. Developments in packaging materials and technologies have made the application of modified atmosphere packaging (MAP) on a larger scale to meat and meat products feasible (Brody, 1989).

Packaging a perishable product in an atmosphere which has been modified so that its composition is other than that of air is termed as MAP (Hintlain and Hotchkiss, 1986). In contrast, controlled atmosphere storage (CAS) involves maintaining a precisely defined atmosphere in the storage chamber by continuous monitoring and compensating for atmospheric changes due to product/microbial respiration or package permeability.

The main purposes of MAP of meat and meat products is two fold: to ensure the (i) microbiological shelf life and (ii) the sensory quality of the product, including the color, odor and palatability. Conversion of meat animals to meat invariably results in contamination of the previously sterile muscle interior with spoilage flora and pathogenic species. The microorganisms causing spoilage often out-compete pathogenic organisms and limit the storage life of the products due to production of undesirable colors and odors. Thus, technologies that provide extension of shelf life of the product, both sensory and microbiological, without compromising their safety are being sought. MAP addresses both these objectives, by using combinations of gases in the product environment that limit or reduce growth of spoilage and pathogenic microflora of meat and stabilize the color of the meat.

Gases Used in MAP of Meat

Carbon dioxide, oxygen and nitrogen are the commonly used gases in MAP, along with trace amounts of carbon monoxide, argon and helium. Each of these gases has specific properties and functions and influences the shelf life (microbial and sensory) of the MAP of meat and meat products. In most cases, combinations of these gases are used to minimize the microbial spoilage and improve the color and other sensory characteristics.

Oxygen: The concentration of oxygen in the package determines the oxidative state of the myoglobin in fresh meats (Fig. 1) and the microbiological shelf life of the product. Traditionally, for fresh meats, oxygen concentrations greater than 5% are required to create the desired bright red color (Holland, 1980), while concentrations less than 2% will result in brown color due to metmyoglobin formation (Seideman and Durland, 1984).
Carbon dioxide: The bacteriostatic and fungistatic properties of carbon dioxide have been widely recognized since the 1920s and was used in shipments of beef, mutton and lamb from Australia and New Zealand to England. Carbon dioxide is highly soluble in water, and its’ solubility increases with decreasing temperature and higher meat pH. Greater than 99% of the gaseous carbon dioxide exists as dissolved gas and less than 1% as carbonic acid (H$_2$CO$_3$), which partially dissociates to give H+, HCO$_3^-$ and CO$_3^{2-}$. Although drop in surface pH of meat due to dissolution of CO$_2$ in meat does not fully explain the bacteriostatic effects of CO$_2$, it does contribute significantly to its antimicrobial activity. At lower product temperatures, the solubility of CO$_2$ is greater, and hence is more effective in retarding microbial growth. Thus, CO$_2$ in the product atmosphere in combination with strict temperature control will improve the microbiological quality of the product.

Although older literature indicates that CO$_2$ concentrations greater than 20% result in undesirable brown color in muscle and bone due to either formation of metmyoglobin or precipitation of sarcoplasmic proteins (Seideman and Durland, 1984), this probably was due to small amounts of residual oxygen.

Nitrogen: Nitrogen is normally used as inert filler, and to prevent package collapse when carbon dioxide dissolves into meat tissue (Lambert et al., 1991). While some studies reported extension of shelf life by nitrogen, others did not indicate any antimicrobial/bacteriostatic activity. The variability in shelf life extensions observed in some of the studies could be due to the residual oxygen concentrations in the MAP products due to incomplete evacuation of air from the packages. Thus, inclusion of nitrogen in MAP atmospheres should provide an advantage in terms of exclusion of oxygen and thus, in preventing aerobic spoilage microflora in the meat products.

Carbon monoxide: CO can be used as part of the MAP gaseous mixtures, ranging from 0.3-0.5 %, for stabilization of fresh meat color. Carbon monoxide (CO) has been approved by FDA (2002) for use as a component of a gas mixture in a MAP system as Generally Recognized as Safe (GRAS) up to 0.4%. Although use CO as part of the gaseous mixture of MAP packaged meats has been limited in the United States due to its toxicological effects, the GRAS status should provide the impetus to its increased use. In Norway, MAP of meats using mixtures of carbon monoxide has an estimated 50-60% of the retail market share. Carbon monoxide extends the lag phase and slows growth rate of E. coli, Achromobacter and P. fluorescens (Gee and Brown, 1981) at concentrations of 25-30%, while P. aeruginosa is unaffected even at these high concentrations. However, the use of carbon monoxide at the low concentrations normally recommended (1%) would have relatively little effect on bacterial growth on fresh meats (Hunt et al., 2002).

<table>
<thead>
<tr>
<th>Organism</th>
<th>O$_2$ Requirement*</th>
<th>Max. specific growth rate (h$^{-1}$)</th>
<th>% Inhibition by high CO$_2$</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>5% CO$_2$</td>
<td>100% CO$_2$</td>
</tr>
<tr>
<td>Spoilage Organisms</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Brochothrix thermophacta</td>
<td>F</td>
<td>0.08</td>
<td>0.06</td>
</tr>
<tr>
<td>Citrobacter freundii</td>
<td>F</td>
<td>0.43</td>
<td>0.28</td>
</tr>
<tr>
<td>Escherichia coli</td>
<td>F</td>
<td>0.40</td>
<td>0.27</td>
</tr>
<tr>
<td>Lactobacillus 173</td>
<td>M</td>
<td>0.13</td>
<td>0.12</td>
</tr>
<tr>
<td>Lactobacillus viridiscens</td>
<td>M</td>
<td>0.13</td>
<td>0.11</td>
</tr>
<tr>
<td>Streptococcus faecalis</td>
<td>F</td>
<td>0.41</td>
<td>0.29</td>
</tr>
<tr>
<td>Foodborne Pathogens</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Aeromonas hydrophila</td>
<td>F</td>
<td>0.34</td>
<td>0.16</td>
</tr>
<tr>
<td>Bacillus cereus</td>
<td>A/F</td>
<td>0.24</td>
<td>0.08</td>
</tr>
<tr>
<td>Clostridium sporogenes</td>
<td>An</td>
<td>0.20</td>
<td>0.12</td>
</tr>
<tr>
<td>Staphylococcus aureus</td>
<td>A/F</td>
<td>0.17</td>
<td>0.09</td>
</tr>
<tr>
<td>Yersinia enterocolitica</td>
<td>F</td>
<td>0.27</td>
<td>0.20</td>
</tr>
<tr>
<td>Yersinia fredericksenii</td>
<td>F</td>
<td>0.25</td>
<td>0.12</td>
</tr>
</tbody>
</table>

Table 1. Anaerobic growth rates of spoilage and pathogenic organisms in the presence of low and high CO2 levels (From Jones, 1989 and Lambert et al., 1991). *A: Aerobe, requires oxygen for growth; An: Anaerobe, inhibited by oxygen; F: Facultative, grows in the presence or absence of oxygen; M: Microaerophile, requires low levels of oxygen.
Combinations of carbon monoxide with other gases such as carbon dioxide to control microbial growth provide an excellent opportunity for meat processors to improve shelf lives of the retail packed fresh meats (Kropf, 1980). With an increasing trend towards case-ready meats, carbon monoxide can provide the means to achieve the retail display life of fresh meats for today’s industry.

**Meat Color**

Meat color is the single greatest appearance factor that determines whether or not a meat cut will be purchased (Kropf, 1980). The color of muscle tissue is determined by the concentration of oxygen (Fig. 1) and the oxidation state of the muscle pigment, myoglobin (Fig. 2). The display life of meat is limited by the time required for oxidation of oxymyoglobin to metmyoglobin, initially on the surface layers of muscle tissue, and reaches proportions of total pigment concentrations that the meat appears dull, and eventually brown (Gill, 1995).

During normal distribution of meat products, the primals and sub-primals are marketed to the retailers vacuum packaged, and the retailer fabricates these into smaller retail cuts, and displays them in over-wrapped packages. The myoglobin, normally will be in deoxy- form under vacuum and is converted to oxy-form during fabrication and display. The oxymyoglobin is gradually oxidized to form metmyoglobin, and the kinetics of the process is dictated by several factors such as the muscle type, rate of postmortem pH decline, packaging film, oxygen consumption, display lighting and temperature and the intrinsic metmyoglobin reducing activity of the muscle (Ledward, 1985).

Exposure of pork and pork products to nitrogen in the gaseous environment does not affect the color, and is similar to vacuum packaged products. Presence of carbon dioxide in the MAP gas mixture has been reported to discolor meat. However, this is likely due to presence of small amounts of residual oxygen in the package as metmyoglobin formation is independent of carbon dioxide concentration (Ordonez and Ledward, 1977). Thus, removal of essentially all of the oxygen from the package is essential to delay or prevent discoloration (Jeremiah, 2001).

Oxygen in the environment is necessary to impart the fresh meat color by formation of oxymyoglobin, which is more resistant to oxidation, compared to the deoxy form. The presence of minimal concentrations of oxygen would suffice for the growth of aerobic spoilage flora of meat, and their growth can be delayed by incorporation of 20 to 30% carbon dioxide in the gas mixtures (Gill, 1995).

Incorporation of carbon monoxide in the gas mixture can provide a stable, cherry red color to the meat by formation of carboxy myoglobin, which is more resistant to oxidation compared to the oxymyoglobin (Sorheim et al., 1997). As discussed earlier, combination of carbon monoxide with other gases can provide the advantages of color stability in addition to microbial control.

**Meat Microbial Spoilage**

Fresh meats provide a rich source of nutrients and support microbial growth (Gill and Newton, 1977). Presence of aerobic conditions (non-limiting in oxygen concentrations) results in growth of mainly aerobic psychrotrophic types, Acinetobacter, Moraxella and Pseudomonas. Growth of Pseudomonas results in production of mal-odors, putrid compounds when the available energy source (glucose) is depleted and the amino acids are utilized for energy requirements (Gill, 1976 and Gill and Newton, 1978).

In the absence of oxygen, the aerobic spoilage types are inhibited and spoilage is primarily due to anaerobic...
bic, aerotolerant lactobacilli. In normal meat (pH<5.8), the lactobacilli predominate and produce delayed acid-dairy flavor to the meat subsequent to growth to very high populations (Seideman and Durland, 1983). However, in high pH (>5.8) meats, the facultative anaerobic types, psychrotrophic Enterobacter, Brochothrix thermospachta, and Alteromonas putrifaciens will predominate. While Brochothrix imparts a sour odor, growth of Enterobacter and Alteromonas will result in putrid, sulfurous odors at higher populations.

The microbial spoilage of fresh meats is dictated by the initial microbial quality of the meats, types of organisms present initially, product storage temperature, time and package conditions, including the gaseous atmospheres in the MAP products. It should be borne in mind that evacuation and incorporation of a particular mix of gases does not assure maintenance of the concentrations indefinitely, and respiration of muscle as well as the surface microflora will modify the atmospheres in the MAP products. Thus, the levels and types of microflora will be dynamic, depending on the prevailing atmospheres and their lengths of time.

**Meat Microbial Safety**

The majority of MAP fresh meats are stored refrigerated, and hence pathogens that can survive and grow under these conditions, such as *Listeria monocytogenes*, *Yersinia enterocolitica*, *Aeromonas hydrophila*, *Bacillus cereus* and *Clostridium botulinum* will be discussed.

*L. monocytogenes* has emerged as a pathogen of concern to the meat industry since the 1990s. The organism is psychrotrophic, facultatively anaerobic, and widely distributed in the environment. In spite of its ability to grow under refrigerated temperatures, the organism does not compete well with the normal spoilage flora of meat (Manu-Tawiah et al., 1993). At high concentrations of carbon dioxide (100%), *L. monocytogenes* does not grow even in high pH fresh meat at < 5°C; however, presence of very low levels of oxygen (5%, Gill and Reichel, 1989) or vacuum packaging and storage at 0, 5 or 10°C does allow its growth. Further, *L. monocytogenes* did not grow in MA (75% CO₂ and 25% N₂) in poultry meat, indicating that a combination of carbon dioxide at high concentrations and low storage temperature can control this pathogen.

*Yersinia enterocolitica* is more prevalent in pork and pork products, is a facultative anaerobe and can grow at refrigeration temperatures (psychrotrophic). *Y. enterocolitica* can grow (at 4°C) on fresh pork chops, and its growth rate is enhanced in anaerobic/CO₂ enriched conditions, and is similar to the spoilage microflora, reaching very high population levels within 35 days of storage. Thus, the presence of this pathogen on fresh meat can result in growth and may reach substantial populations to present a microbiological risk when stored under vacuum or MAP.

*Aeromonas hydrophila* is a psychrotrophic, facultative anaerobe, although rarely isolated from fresh pork and pork products, can pose a microbiological safety risk. *A. hydrophila* can grow on high pH meat in vacuum, while in CO₂ enriched atmospheres, its growth is inhibited (Gill and Reichel, 1989). The fact that the organism can survive in pork stored under N₂ for 10 days at 4°C, but not under carbon dioxide, indicates that the organism is sensitive to carbon dioxide at low temperatures. Storage of pork loin slices at 1°C under MAP increased the shelf life to >16 days, along with control of *A. hydrophila*, while a 7 day shelf life was observed for slices packaged in air (Mano et al., 2000).

*Bacillus cereus* is a spore former, and although the majority of the isolates are mesophilic, some psychrotrophic strains have been reported. Effects of MAP gases on germination and growth of this pathogen in fresh meats have not been investigated.

*Clostridium botulinum* belonging to group II (non-proteolytic) can grow under refrigerated, anaerobic conditions. Use of carbon dioxide concentrations >45% may provide a degree of safety in terms of delaying *C. botulinum* toxigenesis. The incidence of psychrotrophic *C. botulinum* is rare in pork and pork products, and strict temperature control at 1°C can reduce the risk of its growth and toxigenesis.

**Safe Shelf Life Prediction**

In a logical extension of predictive modeling, Labuza et al. (1992) described a method to determine the shelf life of MAP meat products taking into consideration the microbial spoilage, microbiological safety and the visual/sensory deterioration of the food products. During development of a new product or application of a new packaging system for existing product, such as MAP, it is necessary to evaluate the micro-
biological risks scientifically, and determine the safe shelf life of the product.

In MAP products, where higher concentrations of carbon dioxide are used to control spoilage flora, and carbon monoxide is incorporated into the gas mixtures to prevent or reduce the organoleptic/visual spoilage, extreme care must be taken in determining the safe shelf life of the product and strict adherence to the sell-by/use-by dating procedures should be followed to prevent microbiological risks.

Conclusions

Although the use of MAP in meat and meat products has been practiced for almost a century in Australia/New Zealand and Europe, the potential incentives that can be achieved using this technology has not been realized in North America. However, caution should be exercised in determining the optimal combinations of the gas mixtures, with strict adherence to temperature control and comprehensive evaluation of safety of the product should be conducted prior to introducing these products in the market.

References


Information developed for the Pork Information Gateway, a project of the U.S. Pork Center of Excellence supported fully by USDA/Agricultural Research Service, USDA/Cooperative State Research, Education, and Extension Service, Pork Checkoff, NPPC, state pork associations from Iowa, Kentucky, Missouri, Mississippi, Tennessee, Pennsylvania, and Utah, and the Extension Services from several cooperating Land-Grant Institutions including Iowa State University, North Carolina State University, University of Minnesota, University of Illinois, University of Missouri, University of Nebraska, Purdue University, The Ohio State University, South Dakota State University, Kansas State University, Michigan State University, University of Wisconsin, Texas A & M University, Virginia Tech University, University of Tennessee, North Dakota State University, University of Georgia, University of Arkansas, and Colorado State University.